SYNTHETIC BIOTICS AS A VERSATILE PLATFORM TO TREAT INFLAMMATORY BOWEL DISEASE

Bat-Erdene Jugder, Jenny Shu, Analise Reeves, Anna Sokolovska, Jorge Ticas, Chun-Cheih Chao, Jillian Means, JR Gao, Catherine Monahan, David Lubkowicz, Lauren Renaud, Mylene Perreault, Vincent Isabella, and David L. Hava Synlogic Inc., Cambridge, MA, USA

Summary

- Synthetic Biotics are live, non-pathogenic *E. coli* Nissle (EcN) bacteria designed with drug-like properties.
- Our synthetic biology platform allows the engineering of probiotic strains with versatile and disparate modalities that can influence inflammatory pathways in vivo including:
 - Short chain fatty acid and lactate production that can alter inflammatory signaling
 - Aryl hydrocarbon receptor (AHR) metabolites to reduce inflammation and enhance mucosal healing in preclinical IBD models
 - Production and secretion of human proteins, including cytokines
- These innovations are key to the development of gut-targeted therapeutics for inflammation-based pathologies such as Inflammatory Bowel Disease (IBD).

Engineering EcN to produce Short Chain Fatty Acids and Lactate



Fig. 1. A. Schematic of EcN-Lac. The lactate producing strain was engineered by removing the *pta* gene from the acetate producing pathway and introducing a plasmid containing the *E. coli ldhA* gene under the control of an inducible promoter. **B.** Schematic of SFCA metabolic pathways that can be altered for SCFA production. Red Xs denote possible genes modified for a butyrate producing strain. C. In vitro measurement of excess SCFAs produced from engineered EcN strains compared to wild type EcN. Dark blue var denotes EcN-Lac.

D. Mouse study design. 1e10 CFU EcN-Lac were gavaged into wild type mice daily for 3 days prior to anti-CD3 treatment. After 24 hrs, mice were sacrificed and colons were harvested.

E. % of FoxP3⁺ Tregs present in the colon.

F. Gene expression in the colon measured by qPCR and normalized to *Gapdh*.

EcN engineered pathway for IAA production



Fig 2. EcN-IAA strain contains two plasmids: (a) plasmid encoding the enzymatic pathway for tryptophan (Trp) production, *trpE^{fbr}BCDA* (b) plasmid encoding the enzymatic pathway which converts Trp to indole-3-acetic acid (IAA), *trpDH-ipdC-iad1C*. A feedback inhibition resistant DAHP synthase, encoded by *aroG^{fbr}*. Regulation of Trp and IAA production is carried out by P_{fnrs} anaerobicinducible promoter. The *E. coli* Nissle (EcN) chassis contains two key chromosomal deletions: (c) tnaA, encoding tryptophanase, which converts Trp to indole, and (d) trpR, encoding Trp operon repressor, which inhibits production of Trp pathway enzymes.

Engineered EcN-IAA therapy can ameliorate IBD in DSS mouse model





Fig. 3. A. Naïve mice orally dosed with 1E+10. EcN-IAA transits and resides in cecum and colon for at least 6 hrs post-dose. Gut contents were weighed, homogenized and plated for colony formation assay. B. EcN-IAA produces and significantly increases IAA concentration in cecum and colon. Gut contents were weighed and homogenized in 80% methanol and LCMS was used to quantify IAA in gut content. C. EcN-IAA produces bioactive IAA and activates AHR to drive downstream Cyp1a1 expression. D-F. EcN-IAA can reverse diseases progress in DSS induced colitis model by maintaining intestinal barrier. **D.** EcN-IAA was orally dosed before DSS giving and have it colonized by providing selection antibiotic in drinking water for whole experiment. E. CW800 dye was given orally the day before blood harvest. Serum was isolated for signal detection F. Expression of epithelium functional genes in colon. Colon is harvest at day 9, as shown in Fig. 5D. (ANOVA multiple comparison or t test is applied, *, p<0.05; **, p<0.01; ***, p<0.001)

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Development and characterization of EcN-IL-22, an integrated hIL22 Secreting Strain



Fig. 4. A. Schematic of EcN-IL-22. Multiple copies of IL-22 fused to a secretion signal (SS) are integrated into the EcN chromosome. Removal of the *pal* gene results in a Diffusible Outer Membrane (DOM) phenotype for enhanced secretion from EcN. B. ELISA assay comparing the affect of plasmid copy number, ribosome-binding site (RBS) and signaling peptide on hIL-22 secretion normalized to 1e8 CFU/5hrs. hRBS, high affinity RBS C. IL-22 production from integrated strains containing 1, 2, or 4 copies of integrated hIL-22 with the ompA signaling peptide. The 4-copy integrant is designated EcN-IL-22.

hIL22 secreted by EcN induces IL22 dependent gene expression in vivo



Fig. 5. A. EcN-IL-22 is viable and biologically active *in vivo*. EcN-IL-22 numbers decrease with time in naïve mice with Strep in drinking water. **B.** At day 4, high levels of secreted hIL-22 are detected in the colon by ELISA. In colon tissue, *in vivo* target engagement of bacterially-secreted IL-22 is detected as upregulation of the IL-22dependent biomarker, Reg3b.